

Early child growth: how do nutrition and infection interact?

Kathryn G. Dewey and Daniel R. Mayers

Summary of main points

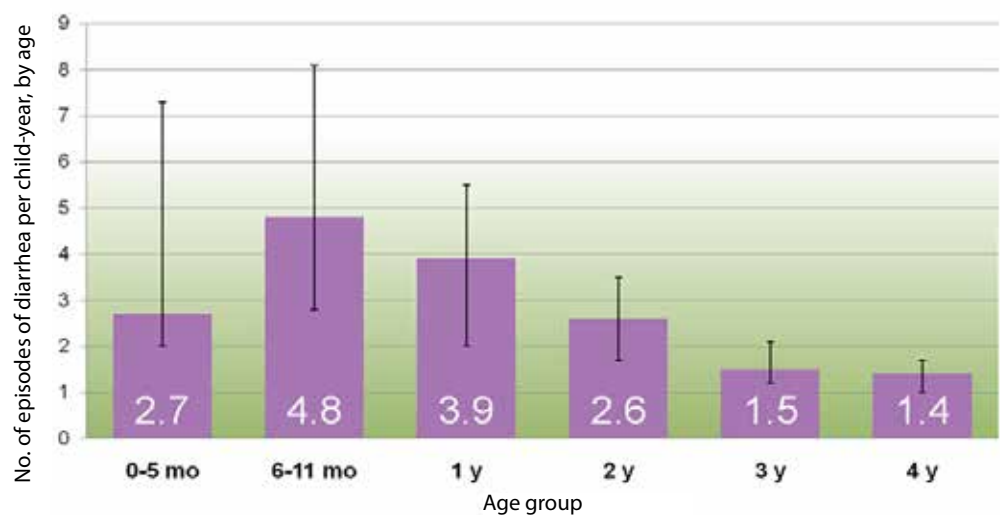
1. Infections are very common in the first 2 years of a child's life.
2. Even when there are no obvious symptoms, physiological conditions associated with infections can impair growth by:
 - Suppressing appetite
 - Impairing absorption of nutrients and increasing nutrient losses
 - Diverting nutrients away from growth
3. There is little direct evidence that nutrition interventions are less effective when infection is common. Further research is needed.
4. Four intervention trials showed that the negative effects of diarrhea on growth can be reduced or eliminated by improved nutrition.
5. Interventions that combine improved nutrition with prevention and control of infections are likely to be most effective for enhancing child growth and development.

This brief was prepared by Kathryn G. Dewey and Daniel R. Mayers. The Insight series of technical briefs addresses the continuum of care for good infant and young child feeding, from initiation of early and exclusive breastfeeding through complementary feeding in later infancy and the second year of life. Alive & Thrive aims to improve feeding practices during this critical period to save lives, prevent malnutrition, and promote optimal growth. The series is developed by the Alive & Thrive team: FHI 360, BRAC, GMMB, IFPRI, Save the Children, UC-Davis, and World Vision.

Infections are very common in the first two years of life. For example, children under two experience an average of 3-5 episodes of diarrhea per year in developing countries (figure 1). In some countries the rate is 6-8 episodes per year. Diarrheal incidence peaks at 6-11 months of age as infants eat increasing amounts of complementary foods that may be contaminated. At this time they begin to crawl and explore their environment, putting them in direct contact with multiple sources of pathogens. During an infection, the immune system requires a broad range of nutrients to mount a defense against the invading organism. It has been hypothesized that nutrition interventions targeting growth may not be effective if infections are prevalent. On the other hand, improved nutrition may strengthen the child's ability to fight infection and reduce the negative effects of infection.

The objective of this brief is to review the available evidence on whether infection diminishes the positive impact of nutrition interventions on child growth and whether improved nutrition limits the negative impact of infections on child growth – i.e., the *interaction* between nutrition and infection. Our purpose is not to examine the direct effect of nutrition on infection, which is a vast topic that goes beyond the scope of this brief. We begin with an overview of the relationship between child growth and the two most common categories of infection: diarrheal and respiratory infections. We then discuss the potentially growth-suppressing impact of subclinical infections and conditions, i.e., the ones that cause no obvious outward symptoms but may have important physiological effects. Next, we examine the evidence on whether there is an interaction between nutrition and

Figure 1: Episodes of diarrhea per year among children under five¹



Source: Bulletin of the WHO 2003;81:197-204
Bars represent the 25th-75th percentiles across 20 countries (1990-2000)

infection with respect to child growth. We conclude with a brief discussion of the programmatic implications.

How strong is the impact of diarrheal and respiratory infections on child growth?

Diarrheal disease has many causes including pathogenic bacteria and other infectious microorganisms. In most cases, exposure to these pathogens occurs through the ingestion of contaminated food and water. Diarrheal illness is generally self limiting, meaning that the infection will run its course and the child will return to normal without requiring specific treatment. However, severe or persistent diarrhea and repeated exposure to pathogens that affect the gut can have serious consequences. Diarrhea robs the child of fluids and certain key nutrients such as zinc and copper.² If these fluids and nutrients are not replaced, the result can be severe dehydration, malnutrition, growth faltering, and death in extreme cases.

It is normal for children to exhibit growth faltering during a bout of diarrhea and to grow more rapidly than usual (“catch-up” growth) after recovery, but the extent of “catch-up” growth may depend on the age of the child, the child’s initial nutritional status, the specific pathogen(s) causing infection, the duration of the infection, and the duration of the “diarrhea-free” interval following infection.^{3,4} For example, children in Peru who were infected with the microorganism *C. parvum* experienced both weight and height growth faltering for several months post-infection followed by periods of “catch-up” growth. Infants took longer to “catch-up” in weight than children infected after 12 months of age, and those who were infected between birth and 5 months of age had a deficit of nearly 1 cm in height one year after infec-

tion, compared to non-infected infants.³ Children who were already stunted (low length-for-age) at the time of infection did not catch up in either weight or height within one year after infection. Those who were not stunted at the time of infection achieved catch-up weight within approximately 3 months and catch-up height within approximately 6 months after infection, compared to their non-infected counterparts.

A high burden of diarrhea in the first two years of life is associated with a much higher risk of stunting (height-for-age < 2 standard deviations below the norm). In a pooled analysis of data from nine studies in five countries (Bangladesh, Brazil, Ghana, Guinea-Bissau, and Peru), 25 percent of stunting at 24 months of age was attributed to having five or more episodes of diarrhea in the first 2 years.⁵ There was a “dose-response” relationship between the cumulative burden of diarrhea (e.g., proportion of days with diarrhea) and the likelihood of being stunted at 24 months of age. Adjusting for socio-economic status did not alter these results.

The impact of respiratory infections on growth is less clear, in part because of a paucity of research on this relationship. The most common types of respiratory infections - mild, upper respiratory infections - are unlikely to have persistent effects in most children. But respiratory infections that include fever are linked with a higher risk of stunting. In a longitudinal study of children in the Philippines followed from birth to 24 months of age, the cumulative impact of febrile respiratory infections on risk of stunting was similar to that of diarrhea.⁶ Fever is one indicator of immune system activation, which (as explained below) can suppress appetite and lead to re-allocation of nutrients away from growth.

What is the role of subclinical infections and related conditions?

An infection is defined as subclinical when there are no obvious signs or symptoms, but there is physiological evidence of abnormality. Young children often test positive for certain infections (e.g., *Helicobacter pylori*, Epstein-Barr virus, cytomegalovirus, mycobacteria, cryptosporidium, and even HIV) without exhibiting clinical symptoms. Many children also carry malaria parasites or gastrointestinal parasites with no outward signs. Subclinical abnormalities of the gastrointestinal tract, presumably caused by frequent exposure to pathogens, are also thought to be common. Even though symptoms are not evident, these subclinical conditions may have a strong, perhaps cumulative effect on metabolic function and growth. Microorganisms in the gut play a critical role in these functions.⁷ The types and relative amounts of different gut bacteria can be affected by the diet.⁸

Environmental Enteropathy

One subclinical condition that is likely to be prevalent in developing countries is environmental enteropathy (EE), also known as tropical enteropathy. This condition often has no outward manifestation but can cause nutrient malabsorption by changing the structure and function of the small intestine. It has been hypothesized that EE causes growth faltering and may decrease the efficacy of nutritional interventions.^{9,10}

EE has been linked to living conditions with poor sanitation and hygiene practices, and is thought to be caused by chronic ingestion of pathogenic microorganisms. Gut exposure to high levels of harmful microorganisms results in a near continuous state of immune system

activation (see next section), which is harmful to the affected individual. Evidence that EE is related to sanitation and hygiene practices is provided by studies of Peace Corps volunteers and U.S. soldiers stationed abroad who developed the condition during their assignments and regained normal intestinal function upon returning home. Research from the 1960s suggested that nearly all residents of the developing world at that time showed some signs of EE.¹¹

Environmental enteropathy is characterized by various small intestinal abnormalities in seemingly healthy individuals. EE can be diagnosed by microscopic examination of an intestinal sample or by laboratory tests for intestinal permeability (sugar ratios present in the urine) or antibodies indicating that bacteria have been able to cross from the intestines into the body (endotoxin-core antibody). In healthy individuals, the surface of the small intestine is covered in millions of tiny, finger-shaped projections called villi. This architecture has evolved to maximize the surface area of the small intestine to facilitate nutrient absorption (figure 2). In a person affected by EE, changes occur in the structure of the small intestine including decreased villous height, sometimes referred to as “flat architecture” (figure 3).

Not all people with EE will experience complete loss of villi; however, regardless of villous architecture, people with EE have increased intestinal inflammation and other structural changes that indicate an elevated immune response.¹² EE is typically associated with a “leaky gut” (increased permeability of the intestinal tract) and impaired ability to prevent pathogens from breaching the intestinal barrier.

The potential pathways by which fecal contamination leads to EE and sub-

sequently to child undernutrition are illustrated in figure 4. Exposure to the causative agents of EE appears to occur very early in life.¹¹ In a comparison of intestinal tissues from stillborn fetuses and young infants in developing countries, the fetal tissues exhibited normal finger-shaped villi, but the tissues from infants showed the flat architecture associated with EE within 3 months after birth. These changes may result in malabsorption of certain nutrients such as vitamin B₁₂ and fats.^{10, 11, 13}

In a study in Peru, children in the worst conditions for sanitation and hygiene experienced 54 percent more diarrheal episodes between birth and 24 months of age and were 1 cm shorter at 24 months than children living in the best conditions. However, the association of water quality and sanitation with height was independent of the association with diarrheal disease. The investigators speculated that constant exposure to harmful bacteria could be causing EE and hindering the children’s ability to effectively absorb and utilize ingested nutrients, regardless of whether it caused diarrhea.¹⁴

EE is probably far more common than overt diarrheal illness in such settings. In a cohort of children in the Gambia, increased intestinal permeability was identified in 76 percent of the 922 samples collected from 119 children between birth and 2 years of age, whereas children were reported to have diarrhea on 7.3 percent of all days during this period.¹⁵ Based on the negative correlation between intestinal permeability and monthly length gain (corrected for age), the investigators calculated that impaired intestinal permeability accounted for 43 percent of linear growth faltering during this period. In a subsequent study¹⁶ in the

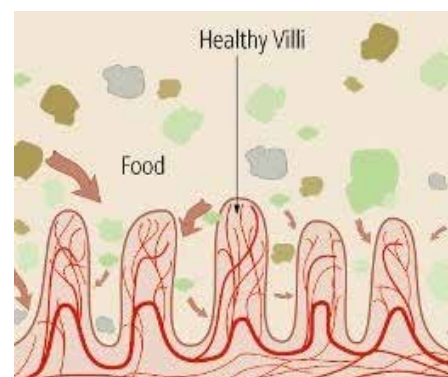


Figure 2 : Normal small intestine with healthy villi

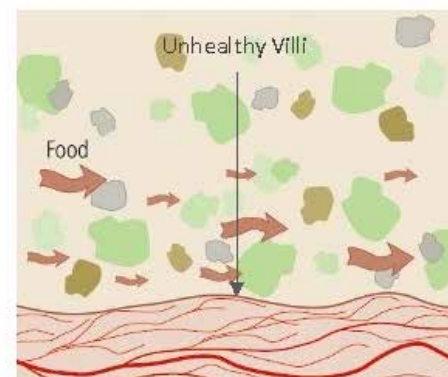
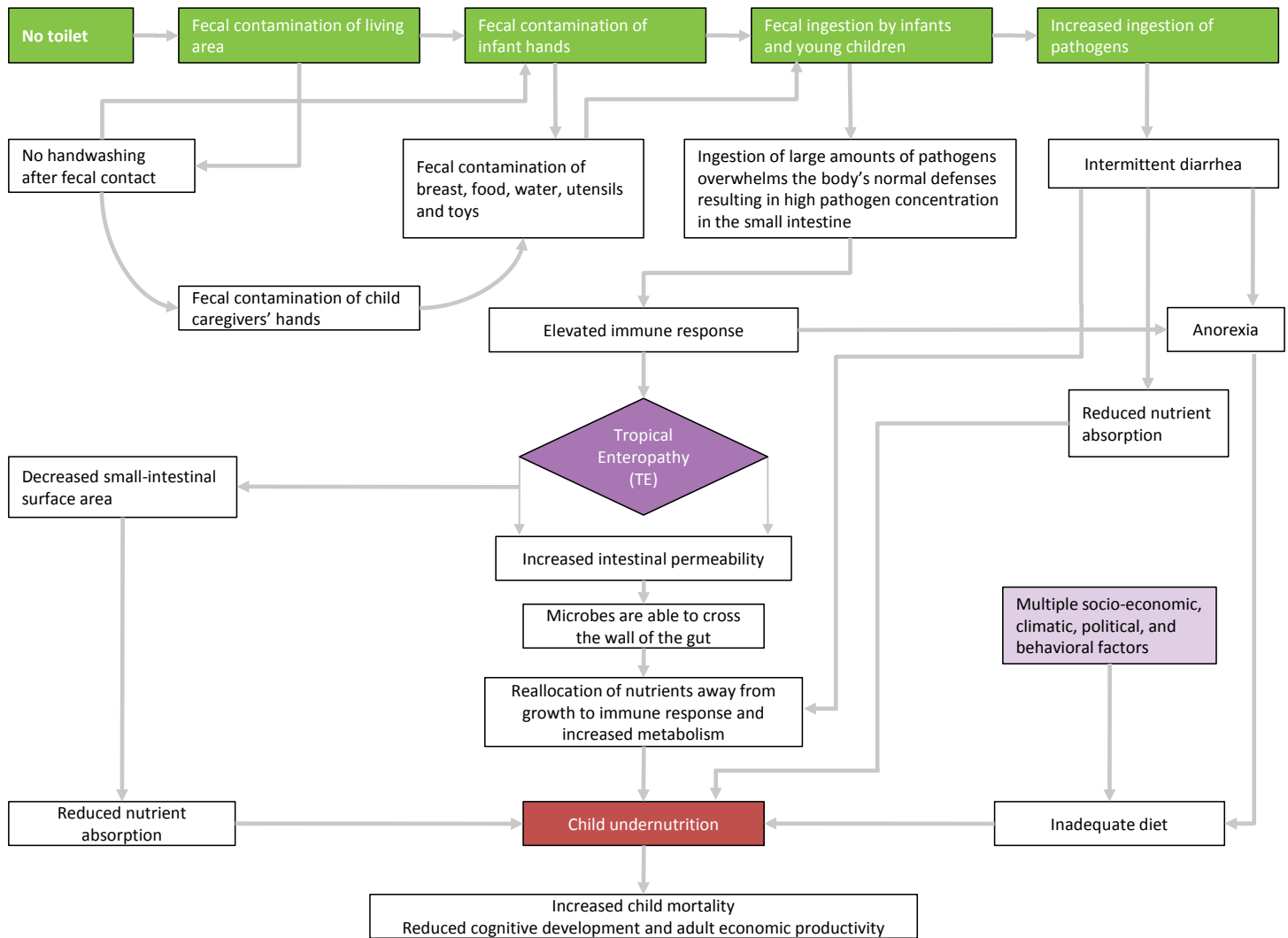


Figure 3: Small intestine with environmental enteropathy characterized by unhealthy villi

Source: <http://duncanmarasanitation.blogspot.com/2009/09/tropical-enteropathy-3.html>

same population, markers of intestinal function were normal at 2 months of age but deviated sharply from the norms by 15 months of age. The combined effects of three different markers of intestinal function were calculated to explain 56 percent of linear growth faltering. Because these were observational studies and the investigators did not control for potentially confounding variables, it is difficult to ascribe cause and effect, but the results point to the high prevalence of EE as a key risk factor for stunting in this population.

Figure 4: Pathways by which fecal contamination leads to environmental enteropathy and child undernutrition
(Adapted from Humphrey 2009)



Immune Activation, Cytokines, and Appetite

In response to infection, the immune system becomes activated and produces specific immune cells and cytokines in large amounts to combat the invading organism. Cytokines are protein molecules that assist in fighting infection. They are beneficial in the short term; however, a chronic condition – like EE – can lead to continuously high levels of cytokines, which can cause negative metabolic consequences and suppress appetite.¹⁷

Reports of “poor appetite” by caregivers of children under two in developing countries are common and may account for a substantial proportion of low energy intakes in this age group.¹⁸ Appetite is controlled by a group of chemicals called satiety hormones. Two important hormones involved in appetite regulation are ghrelin and leptin. Ghrelin stimulates food intake; leptin suppresses food intake. During infection, elevated levels of cytokines can lead to increased blood leptin concentrations and diminished appetite.¹⁹ This effect has been verified in

cases of severe systemic infection such as neonatal blood infections.²⁰

Immune system activation also lowers circulating levels of certain nutrients, in particular vitamin A and zinc, and increases iron retention in the liver, which restricts the availability of iron to other tissues in the body. These effects are probably part of an adaptive response to withhold key nutrients from invading pathogens, but they can result in inadequate availability of certain nutrients to support growth, even if intake is

adequate, during the period of metabolic disturbance. Among children 6-20 months of age in Zambia, blood markers of inflammation (usually associated with infection) were negatively related to growth in length during the subsequent 3 months.²¹

The “Dirty Chicken” Experiment

Studies of how sanitation affects the growth of newly hatched chickens provide clues that may be relevant to growth faltering of children in developing countries.²² A classic experiment conducted in 1992, called the “dirty chicken” study, involved raising chicks in either steam-cleaned or unclean cages in close proximity with their own feces. In each of the two living environments, the chicks were either administered an antibiotic cocktail or no antibiotics. Unsurprisingly, living in close proximity to feces in the poor sanitation environment caused the chicks raised without antibiotics to experience decreased rates of weight gain, decreased efficiency of food utilization, and increased levels of the cytokine plasma interleukin 1.

However, chicks raised in poor sanitary conditions and given antibiotics grew just as well and had the same low levels of circulating cytokines as chicks raised in steam-cleaned cages. The investigators concluded that the administration of antibiotics facilitated growth by preventing the immunologic stress and associated metabolic changes brought about by chronic exposure to feces.

Researchers have attempted to treat EE in human children with antibiotics but with little success. In fact, in one study provision of antibiotics led to an increased incidence of diarrhea, perhaps due to a negative effect of antibiot-

ics on the “good” bacteria in the gut.²³ Short-term antibiotic therapy may fail because of re-exposure to fecal bacteria soon after treatment. Without improved sanitation and hygiene practices, a single course of antibiotics is unlikely to reverse EE, which may take months to resolve especially if there is repeated exposure to pathogens.^{24, 25}

While prevention of infection-related growth faltering with antibiotics in humans may not be feasible, the “Dirty Chicken” experiment indicates that living in poor sanitary conditions can cause growth faltering and implies that decreasing the burden of infection – including subclinical infection – may significantly improve growth outcomes.

What is the interaction between nutrition and infection?

Does infection make nutrition interventions less effective?

During infection, energy and other nutrients are diverted towards the immune response and away from growth. After all, survival is more important than continuing to grow, so growth faltering during infections may be an adaptive mechanism. However, repeated episodes of infection or persistent subclinical infection may put the child in a near-constant state of growth suppression. Does this mean that nutritional interventions for populations with high exposure to infections will be unsuccessful at improving child growth?

Evidence on this question is scant. In Indonesia, the effect of high-dose vitamin A supplements on linear growth in preschool children (6-48 months of age) was dependent on the burden of respiratory infection.²⁶ In children with a low

burden of respiratory infection, especially those with low vitamin A intake, linear growth improved after vitamin A supplementation. In children with a high burden of respiratory infection, there was little or no impact of vitamin A supplementation on growth regardless of vitamin A intake.

One potential explanation is that supplemental vitamin A is not well absorbed during an acute infection and a large proportion is excreted in the urine, rendering the high-dose supplement much less effective at improving vitamin A status if it is administered when the child is ill. Another potential explanation is that fever during respiratory infections causes metabolic changes that reduce circulating levels of vitamin A and make it less available to tissues to support growth. Regardless of the mechanism for the effect, the investigators concluded that coupling vitamin A supplementation programs with efforts to reduce respiratory infections would increase the likelihood of a positive impact on growth.

Apart from the single study described above, there is little direct evidence that the impact of nutrition interventions is blunted when infections are common. Further research on this question is needed.

Does improved nutrition reduce the negative impact of infection?

The contrasting hypothesis is that improved nutrition can lessen or even eliminate the negative impact of infections on growth. The potential mechanisms by which improved nutrition could reduce the impact of infections on growth are shown in box 1. These mechanisms include a) strengthening

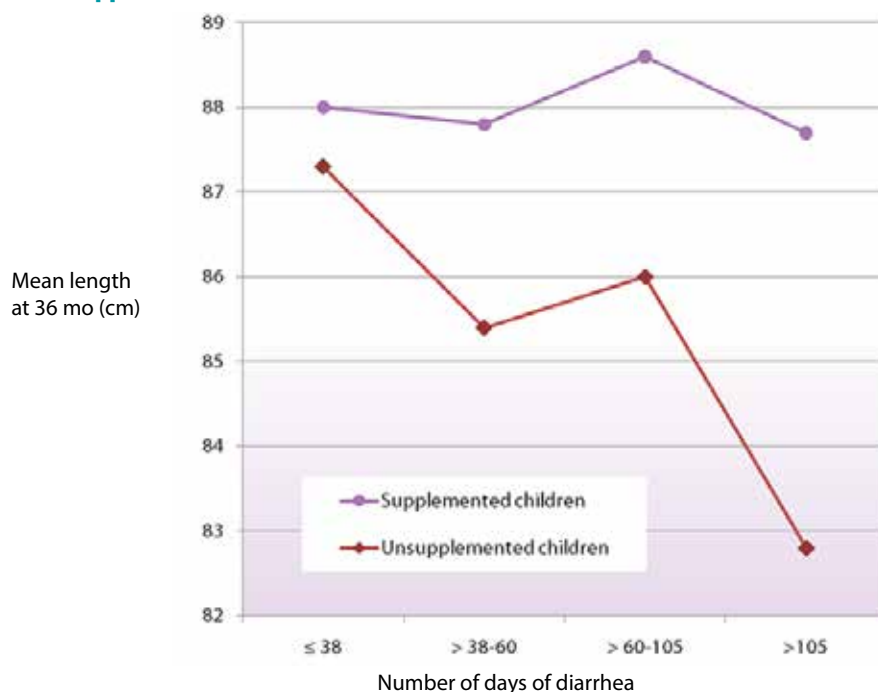
Box 1. Improved nutrition may reduce the negative impact of infections on growth by:

1. Strengthening the child's immune system, thereby reducing the severity and duration of infections and their impact on growth
2. Providing extra amounts of nutrients to compensate for those that are not well absorbed during infection, lost during diarrhea, reallocated elsewhere in the body due to immune system activation, or consumed in lower amounts than usual because of reduced appetite during infection
3. Providing the required amounts of nutrients for catch-up growth following infection, particularly the nutrients that are needed to build lean body tissue such as protein, potassium, magnesium, phosphorus, zinc, and sodium
4. Preventing poor appetite caused by micronutrient deficiencies, thereby facilitating catch-up growth
5. Favoring the growth of beneficial bacteria in the gut that enhance gut function and immune defenses

the immune system, b) compensating for malabsorption, reallocation, or losses of key nutrients, c) allowing for catch-up growth following infection, d) enhancing appetite, and e) favoring the growth of beneficial gut microorganisms. Four nutrition intervention trials among preschool children in Colombia, Guatemala, Tanzania, and South Africa indicate that provision of macro- and/or micronutrients can limit the negative effects of diarrhea on child growth.

Randomized food supplementation study in Colombia, 1973-1980. Families were eligible for food supplementation

Figure 5: Relationship between length at 36 months of age and number of days of diarrhea during the first three years of life among supplemented and unsupplemented children in Colombia²⁸



if the mother was in the first or second trimester of pregnancy and at least half of her preschool-aged children were underweight.²⁷ All household members in the intervention group received protein-enriched food that included powdered skim milk. Children in all groups were supplemented with iron and vitamin A. For the analysis of child stunting related to diarrhea, the investigators used data for 241 children followed for the first 3 years of life for whom they had complete morbidity data and measurement of height (length) at 36 months of age (148 unsupplemented children and 140 children who were supplemented from the sixth month of pregnancy up to 36 months).

Diarrhea was very common in the Colombia study. The number of episodes from birth to 36 months was 18 in the unsupplemented group and 16 in the supplemented group (not significantly

different). In the unsupplemented children, height at 36 months was strongly inversely associated with the number of days ill with diarrhea (-0.03 cm for each day of illness, $p < 0.001$). In the supplemented children, there was no relationship between diarrheal illness and height at 36 months (see figure 5; $p < 0.001$ for the interaction).²⁸ The positive impact of supplementation on height (overall, approximately 3 cm) was greatest in children with the highest burden of diarrhea (nearly 5 cm). The investigators concluded that nutritional supplementation eliminated the negative impact of diarrheal disease on child growth. They speculated that improved nutrient intake during and/or after illness episodes facilitated catch-up growth.

Supplementary feeding intervention in Guatemala, 1969-1977. A large supplementary feeding trial targeting pregnant and lactating women and their children

from birth to 7 years of age was conducted in two sets of two matched villages. One village in each set was randomly selected to receive either a high-protein, high-energy supplement called 'Atole' or a non-protein low-energy supplement called 'Fresco', both fortified with several micronutrients.^{29, 30}

Among children 3-36 months of age who received Fresco, there was a significant negative relationship between the percentage of time with diarrhea and length gain. By contrast, among children who received Atole, there was no significant relationship between diarrhea prevalence and length gain ($p < 0.05$ for interaction effect).²⁸ These findings were similar to those from the Colombia trial.

Vitamin A supplementation in Tanzania, 1993-1997. In this study, 687 children 6 to 60 months of age who had been admitted to the hospital with pneumonia were randomly assigned to high-dose capsules of vitamin A or placebo while hospitalized and again 4 and 8 months after discharge.³¹ There was no significant effect of vitamin A supplementation on growth for otherwise healthy children, but in children with persistent diarrhea during the follow-up period, vitamin A eliminated the risk of stunting usually associated with this condition. Specifically, in the placebo group, the risk of stunting (adjusted for potential confounders) was 3.7 times higher in children with persistent diarrhea than in those without persistent diarrhea. In the vitamin A group, there was no risk of stunting associated with persistent diarrhea ($p=0.015$ for interaction effect). Children with persistent diarrhea may have lower levels of circulating vitamin A than children with acute or no diarrhea, so the vitamin A supplements may have compensated for this phenomenon and allowed for catch-up growth in children recover-

ing from persistent diarrhea. Although vitamin A is generally not considered a key growth-limiting nutrient,³² ensuring adequate vitamin A may facilitate growth by restoring other physiological functions that must be normalized to permit rapid gain in lean tissue.

Micronutrient supplementation in South Africa, 2003-2006. In this study, 373 infants in three cohorts (32 HIV-infected children, 154 HIV-uninfected children born to HIV-infected mothers, and 187 uninfected children born to uninfected mothers) were randomly assigned at 6 months of age to receive daily micronutrient supplementation for 18 months with either vitamin A, vitamin A plus zinc, or multiple micronutrients that included vitamin A and zinc.³³ The study showed no overall impact on growth of zinc or multiple micronutrients compared to vitamin A alone (though there was a positive impact of multiple micronutrients on child length in those who were already stunted at enrollment). In the two cohorts of HIV-uninfected children, the addition of zinc or multiple micronutrients to vitamin A reduced the impact of repeated diarrhea episodes on linear growth. This was most evident among children who had more than six episodes of diarrhea per year ($n=34$). In this sub-group, infants who received only vitamin A exhibited a decline of 0.6 Z-scores in length-for-age between 6 and 24 months of age, but those who received multiple micronutrients showed no decline in length-for-age during the same interval ($p=0.06$ within this subgroup of 34). The investigators suggested that the progressive stunting usually observed in children with repeated episodes of diarrhea may be related to deficiencies of certain micronutrients, and could be prevented by adequate intake of those micronutrients.

These four studies all show that the negative effects of diarrhea on growth can be offset by nutrition interventions, at least in these particular situations. However, as mentioned above, clinical symptoms of diarrhea may be just the "tip of the iceberg" when it comes to gastrointestinal conditions that can affect growth. EE may be much more prevalent than diarrhea. Whether nutrition interventions can reduce or eliminate the growth-suppressing impact of EE is unknown.

In adults, however, there is some evidence that multiple micronutrients may partially reverse the impact of EE on gut function. In a study of intestinal impairment in Zambia, 500 adults (with or without HIV infection) were randomly assigned to receive multiple micronutrients or placebo for 2 years. Micronutrients had no impact on markers of intestinal permeability, but there was a significant reduction in one of the markers reflecting bacterial movement across the intestinal wall.³⁴ This suggests an improvement in gut integrity or immune function, but further research, particularly in children, is needed.

To date, there is almost no information on whether improved nutrition can reduce the impact of infections other than gastrointestinal infections, such as respiratory illnesses, and malaria on child growth, though the Tanzania study described above showed that vitamin A supplements were more effective for improving growth in children infected with malaria or HIV than in non-infected children.

Some nutrients, such as iron, have the potential to increase the risk of infection, or mortality due to infection, and may interfere with linear growth.^{35, 36} The mode of administration, such as supplementation vs. fortification, and the initial iron

status of the individual are key factors to consider when evaluating whether nutrition interventions that include iron are likely to reduce or exacerbate the influence of infection on growth.

Conclusions and programmatic implications

Infections play a major role in preventing children in developing countries from reaching their growth potential. A high burden of diarrheal disease is a key risk factor for stunting, and other types of infections also contribute to growth faltering although their impact is not as well documented. However, the view that “disease rather than diet” is the main cause of growth impairment¹⁶ ignores the important interaction between infection and nutrition. To date, the limited evidence available suggests that nutrition interventions can substantially reduce or even eliminate the negative effect of diarrheal disease on child growth. This is encouraging, but it should be recognized that subclinical conditions such as EE may account for a large proportion of growth faltering, and it is not yet known whether improved nutrition can prevent or reverse the deleterious effects of EE (or growth faltering associated with infections other than diarrheal disease).

At present, evidence is insufficient to conclude that high rates of infection make nutrition interventions ineffective for improving child growth. Only one study was found that supported this hypothesis.²⁶ In this study a high burden of respiratory infection limited the potential for vitamin A supplementation to improve growth. Clearly, further research on this issue is needed, but a single study involving a single micronutrient does not warrant holding back on

Box 2. Nutrition and infection prevention and control interventions to improve child growth

- Promote handwashing with soap and water
- Improve sanitation and water quality
- Promote exclusive breastfeeding for the first 6 months and continued breastfeeding thereafter
- Promote appropriate complementary feeding practices including feeding during and after illness and safe preparation and storage of complementary foods
- Step up efforts to prevent and treat respiratory illnesses and other infections such as malaria

efforts to improve nutrition in populations where infections are prevalent.

Nonetheless, combining improved nutrition with efforts to prevent and control infections will likely be the most effective approach for optimizing child growth and development. This question was explored many years ago in the Narangwal Nutrition Experiment conducted between 1969 and 1973 in Punjab, India.³⁷ In that project, ten villages were selected in clusters of two to three villages to receive a package of services that included either nutrition care (NUT), health care focused on infection control (HC), integrated services including both nutrition and health care (NUTHC), or standard care (Control - symptomatic health care on demand only). NUT services included growth monitoring, food supplementation (initially only for malnourished children, but later made available to all children), and nutrition education. Health care services included curative and preventive care for common illnesses, immunizations, and hygiene education. The target group was children under 3 years of age. At 36 months of age, children in the NUT

or NUTHC villages were 1.3 cm taller than children in control villages, with no significant difference between NUT and NUTHC villages. Children in HC villages were taller than those in control villages, but not as tall as children in the NUT or NUTHC villages. Thus, in this setting the combination of nutrition and health care did not produce a greater improvement in growth than nutrition care alone. However, the psychomotor development scores of children in the NUTHC villages generally exceeded the summed separate effects of NUT and HC, suggesting a synergistic effect on those outcomes. Apart from the Narangwal Experiment, very little information exists on whether providing infection control together with direct nutrition interventions has an additive or synergistic effect on child growth or other key outcomes.

Key components of infection control are effective promotion of handwashing with soap and water and improvements in sanitation and water quality, which can significantly decrease diarrheal disease. In a recent meta-analysis,³⁸ handwashing was linked to a 48 percent risk reduction of diarrhea across study designs and pathogens. A substantial positive effect was also found for both water quality and sanitation improvements – 17 percent and 36 percent risk reductions, respectively. Access to and utilization of toilets is a high priority,¹⁰ yet an estimated 2.6 billion people globally live without basic toilets to dispose of feces.³⁹

An essential element of combined approaches is the promotion of breastfeeding for at least two years (exclusively for the first 6 months, and continued breastfeeding in combination with nutritious complementary foods thereafter), which has the dual benefit of reducing infection and improving nutrition. Other

key practices, highlighted in the *Guiding Principles for Complementary Feeding of the Breastfed Child*,⁴⁰ are feeding during and after illness to sustain adequate nutrient intake and promote catch-up growth, and safe preparation and storage of complementary foods to reduce food-borne illnesses.

Although many nutrition programs already include hygiene messages, simply increasing knowledge and awareness about behaviors such as handwashing is not enough. Sustainable changes in behaviors are more difficult to achieve because of factors such as lack of access to clean water, long distance from water sources, the cost of hygiene products, and poor design of educational interventions that do not take into account cultural beliefs or craft messages tailored to the needs and values of the target audience.⁴¹⁻⁴⁴ The difficulty of improving

household hygiene and sanitation practices, outside of intensive efficacy trials, is well recognized by researchers.^{42, 43, 45-47} Innovative strategies have been suggested that focus on emotional motivations for behavior change and engagement of professional consumer and market research agencies, rather than relying solely on knowledge-based approaches.^{41, 45}

Research is needed on the efficacy and effectiveness of approaches that combine nutrition interventions with multiple strategies for prevention and control of infections, including hygiene education, improvements in water quality and sanitation, and measures to prevent and treat respiratory illness and other infections such as malaria. Development and evaluation of integrated cost-effective programs designed to tackle these multiple objectives should be a high priority.

Acknowledgments

The authors, Kathryn G. Dewey and Daniel R. Mayers, gratefully acknowledge the input of the following reviewers of an earlier draft of this brief: Mary Arimond, William Checkley, Sandra Huffman, Jean Humphrey, Steve Luby, Chessa Lutter, Luann Martin, Ellen Piwoz, Andrew Prentice, and Christine Stewart. We also thank Binetti Vitta for her assistance in the preparation of this brief.

Alive & Thrive, launched with a grant from the Bill & Melinda Gates Foundation, is an initiative to improve infant and young child feeding in Bangladesh, Ethiopia, and Viet Nam and inform policies and programs around the world.

For more information visit our website:
www.aliveandthrive.org

References

- Margaret K, Bern C, Guerrant R. The global burden of diarrhoeal disease, as estimated from studies published between 1992 and 2000. *Bull World Health Organ.* 2003; 81(3):197-204.
- Castillo-Duran C, Vial P, Uauy R. Trace mineral balance during acute diarrhea in infants. *J Pediatr.* 1988; 13(3):452-457.
- Checkley W, Epstein LD, Gilman RH, Black RE, Cabrera L, Sterling CR. Effects of *Cryptosporidium parvum* infection in Peruvian children: Growth faltering and subsequent catch-up growth. *Am J Epidemiol.* 1998; 148(5):497-506.
- Wierzba TF, El-Yazeed RA, Savarino SJ, Mourad AS, Rao M, Baddour M, El-Deen AN, Naficy AB, Clemens JD. The interrelationship of malnutrition and diarrhea in a periurban area outside Alexandria, Egypt. *J Pediatr Gastr Nutr.* 2001; 32(2):189-196.
- Checkley W, Buckley G, Gilman RH, Assis AMO, Guerrant RL, Morris SS, Mølbak K, Valentiner-Branth P, Lanata CF, Black RE. Multi-country analysis of the effects of diarrhoea on childhood stunting. *Int J Epidemiol.* 2008; 37(4):816-830.
- Adair LS, Guilkey DK. Age-specific determinants of stunting in Filipino children. *J Nutr.* 1997; 127(2):314-320.
- Preidis GA, Hill C, Guerrant RL, Ramakrishna BS, Tannock GW, Versalovic J. Probiotics, enteric and diarrheal diseases, and global health. *Gastroenterology.* 2010; 140(1):8-14.e19.
- De Filippo C, Cavalieri D, Di Paola M, Ramazzotti M, Poullet JB, Massart S, Collini S, Pieraccini G, Lionetti P. Impact of diet in shaping gut microbiota revealed by a comparative study in children from Europe and rural Africa. *P Natl Acad Sci.* 2010; 107(33):14691-14696.
- Goto R, Mascie-Taylor CGN, Lunn PG. Impact of intestinal permeability, inflammation status and parasitic infections on infant growth faltering in rural Bangladesh. *Brit J Nutr.* 2009;101(10):1509-1516.
- Humphrey JH. Child undernutrition, tropical enteropathy, toilets, and handwashing. *Lancet.* 2009; 374(9694):1032-1035.
- Haghighi P, Wolf PL, Durie P. Tropical sprue and subclinical enteropathy: A vision for the nineties. *Crit Rev Cl Lab Sci.* 1997; 34(4):313-341.
- Sullivan P, Mirakian R, Hill S, Milla P, Neale G, Marsh M. Chronic diarrhea and malnutrition—histology of the small intestinal lesion. *J Pediatr Gastr Nutr.* 1991; 12(2):195-203.
- Ramakrishna BS, Venkataraman S, Mukhopadhyaya A. Tropical malabsorption. *Postgrad Med J.* 2006; 82(974):779-787.
- Checkley W, Gilman RH, Black RE, Epstein LD, Cabrera L, Sterling CR, Moulton LH. Effect of water and sanitation on childhood health in a poor Peruvian peri-urban community. *Lancet.* 2004; 363(9403):112-118.
- Lunn PG, Northrop-Clewes CA, Downes RM. Intestinal permeability, mucosal injury, and growth faltering in Gambian infants. *Lancet.* 1991; 338(8772):907-910.

16. Campbell DI, Elia M, Lunn PG. Growth faltering in rural Gambian infants is associated with impaired small intestinal barrier function, leading to endotoxemia and systemic inflammation. *J Nutr*. 2003; 133(5):1332-1338.
17. Wong S, Pinkney J. Role of cytokines in regulating feeding behaviour. *Curr Drug Targets*. 2004; 5(3):251-263.
18. Brown KH, Peerson JM, Lopez de Romana G, de Kanashiro HC, Black RE. Validity and epidemiology of reported poor appetite among Peruvian infants from a low-income, periurban community. *Am J Clin Nutr*. 1995; 61(1):26-32.
19. Somech R, Reif S, Golander A, Spirer Z. Leptin and C-reactive protein levels correlate during minor infection in children. *IMAJ*. 2007; 9:76-79.
20. Orbak Z, Ertekin V, Akçay F, Ozkan B, Ors R. Serum leptin levels in neonatal bacterial septicemia. *J Pediatr Endocr Metab*. 2003; 16(5):727-731.
21. Hautvast JLA, Tolboom JJM, Willems JL, Mwela CM, Monnens LAH. Consequences of infections for three-month length increment in young children in rural Zambia. *Acta Paediatrica*. 2000; 89(3):296-301.
22. Roura E, Homedes J, Klasing KC. Prevention of immunologic stress contributes to the growth-permitting ability of dietary antibiotics in chicks. *J Nutr*. 1992; 122(12):2383-2390.
23. Trehan I, Shulman RJ, Ou C-N, Maleta K, Manary MJ. A randomized, double-blind, placebo-controlled trial of rifaximin, a nonabsorbable antibiotic, in the treatment of tropical enteropathy. *Am J Gastroenterol*. 2009; 104(9):2326-2333.
24. Thabane M, Marshall J. Post-infectious irritable bowel syndrome. *World J Gastroenterol*. 2009; 15(29):3591-3596.
25. Kirkpatrick BD, Noel F, Rouzier PD, Powell JL, Pape JW, Bois G, Alston WK, Larsson CJ, Tenney K, Ventrone C, et al. Childhood cryptosporidiosis is associated with a persistent systemic inflammatory response. *Clin Infect Dis*. 2006; 43(5):604-608.
26. Hadi H, Dibley MJ, West KP. Complex interactions with infection and diet may explain seasonal growth responses to vitamin A in preschool aged Indonesian children. *Eur J Clin Nutr*. 2003; 58(7):990-999.
27. Lutter CK, Mora JO, Habicht JB, Rasmussen KM, Robson DS, Sellers SG, Super CM, Herrera MG. Nutritional supplementation: effects on child stunting because of diarrhea. *Am J Clin Nutr*. 1989; 50(1):1-8.
28. Lutter C, Habicht J, Rivera J, Martorell R. The relationship between energy intake and diarrhoeal disease in the effects on child growth: biological model, evidence and implications for public health policy. *Food Nutr Bull*. 1992; 14(1):36-42.
29. Ramirez-Zea M, Melgar P, Rivera JA. INCAP oriente longitudinal study: 40 years of history and legacy. *J Nutr*. 2010; 140(2):397-401.
30. Martorell R, Habicht JB, Rivera JA. History and design of the INCAP longitudinal study (1969-77) and its follow-up (1988-89). *J Nutr*. 1995; 125 (4 Suppl):1027S-1041S.
31. Villamor E, Mbise R, Spiegelman D, Hertzmark E, Fataki M, Peterson KE, Ndossi G, Fawzi WW. Vitamin A supplements ameliorate the adverse effect of HIV-1, malaria, and diarrheal infections on child growth. *Pediatrics*. 2002; 109(1):e6.
32. Golden MH. Proposed recommended nutrient densities for moderately malnourished children. *Food Nutr Bull*. 2009; 30(3 Suppl):S267-342.
33. Chhagan M, Van den Broeck J, Luabeya K-K, Mpontshane N, Tomkins A, Bennis M. Effect on longitudinal growth and anemia of zinc or multiple micronutrients added to vitamin A: a randomized controlled trial in children aged 6-24 months. *BMC Public Health*. 2010; 10(1):145.
34. Kelly P, Shawa T, Mwanamakondo S, Soko R, Smith G, Barclay GR, Sanderson I. Gastric and intestinal barrier impairment in tropical enteropathy and HIV: limited impact of micronutrient supplementation during a randomised controlled trial. *BMC Gastroenterol*. 2010; 10(1):72.
35. Hurrell R. Iron and malaria: absorption, efficacy and safety. *Int J Vitam Nutr Res*. 2010; 80(4-5):279-292.
36. Dewey KG, Domelloef M, Cohen RJ, Landa Rivera L, Hernell O, Loennerdal B. Iron supplementation affects growth and morbidity of breast-fed infants: results of a randomized trial in Sweden and Honduras. *J Nutr*. 2002; 132(11):3249-3255.
37. Kiemann A, Taylor C, DeSweemer C, Parker R, Chernichovsky D, Reinke W, Uberoi I, Kakar D, Masih N, Sarma R. Child and maternal health services in rural India: the Narangwal experiment. *Integrated Nutrition and Health Care*, The Johns Hopkins University Press, Baltimore, A World Bank Research Publication. 1983; Vol 1.
38. Cairncross S, Hunt C, Boisson S, Bostoen K, Curtis V, Fung ICH, Schmidt W-P. Water, sanitation and hygiene for the prevention of diarrhoea. *Int J Epidemiol*. 2010; 39(suppl 1):i193-i205.
39. Coombes R. Toiling for toilets. *BMJ*. 2010; 341.
40. Dewey KG, Lutter CK. Guiding principles for complementary feeding of the breast-fed child. Washington, DC: PAHO/WHO; 2003.
41. Biran A, Schmidt W-P, Wright R, Jones T, Seshadri M, Isaac P, Nathan NA, Hall P, McKenna J, Granger S, et al. The effect of a soap promotion and hygiene education campaign on handwashing behaviour in rural India: a cluster randomised trial. *Trop Med Int Health*. 2009; 14(10):1303-1314.
42. Arnold B, Arana B, Mäusezahl D, Hubbard A, Colford JM. Evaluation of a pre-existing, 3-year household water treatment and handwashing intervention in rural Guatemala. *Int J Epidemiol*. 2009; 38(6):1651-1661.
43. Luby SP, Mendoza C, Keswick BH, Chiller TM, Hoekstra RM. Difficulties in bringing point-of-use water treatment to scale in rural Guatemala. *Am J Trop Med Hyg*. 2008; 78(3):382-387.
44. Aunger R, Schmidt W-P, Ranpura A, Coombes Y, Maina PM, Matiko CN, Curtis V. Three kinds of psychological determinants for hand-washing behaviour in Kenya. *Soc Sci Med*. 2009; 70(3):383-391.
45. Curtis V. Talking dirty: how to save a million lives. *Int J Environ Heal R*. 2003; 13(1 suppl 1):73 - 79.
46. Scott BE, Schmidt WP, Aunger R, Garbrah-Aidoo N, Animashaun R. Marketing hygiene behaviours: the impact of different communication channels on reported handwashing behaviour of women in Ghana. *Health Educ Res*. 2008; 23(3):392-401.
47. Luby SP, Agboatwalla M, Bowen A, Kenah E, Sharker Y, Hoekstra RM. Difficulties in maintaining improved handwashing behavior, Karachi, Pakistan. *Am J Trop Med Hyg*. 2009; 81(1):140-145.



www.aliveandthrive.org

Headquarters Office

1825 Connecticut Avenue, NW

Washington, DC 20009

United States

Tel: (202) 884-8000

Fax: (202) 464-3966

Email: aliveandthrive@fhi360.org